

#### **ThermoFisher** SCIENTIFIC

## **Cell Health & Functional Assays for Flow Cytometry**

#### Life Sciences Solutions: Part of Thermo Fisher Scientific





#### Your Thermo Fisher Flow Team

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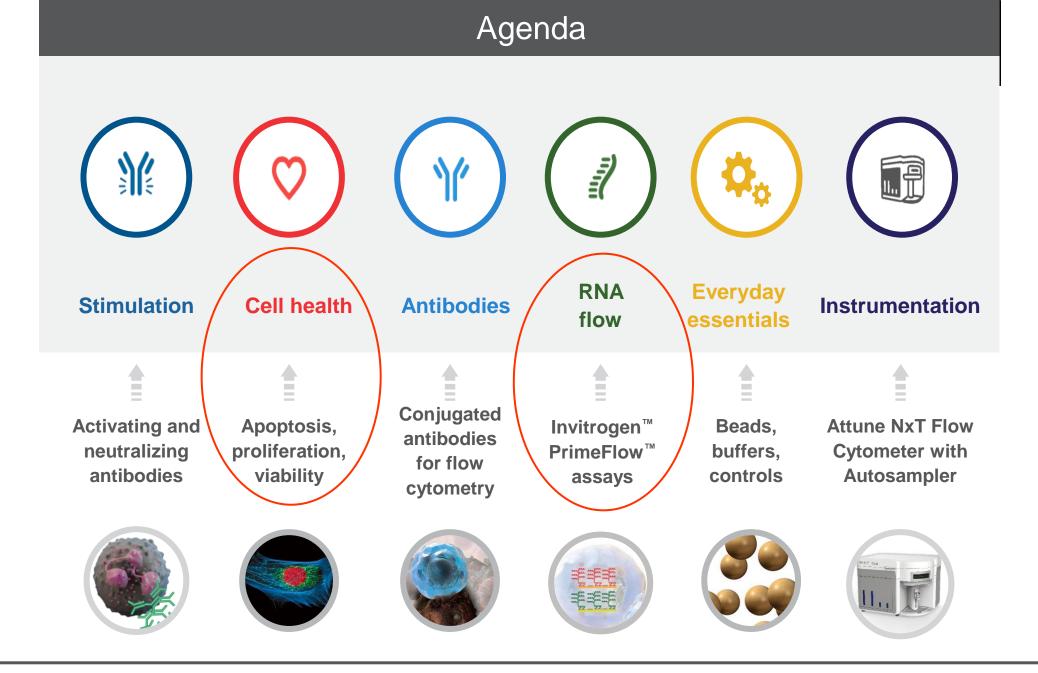
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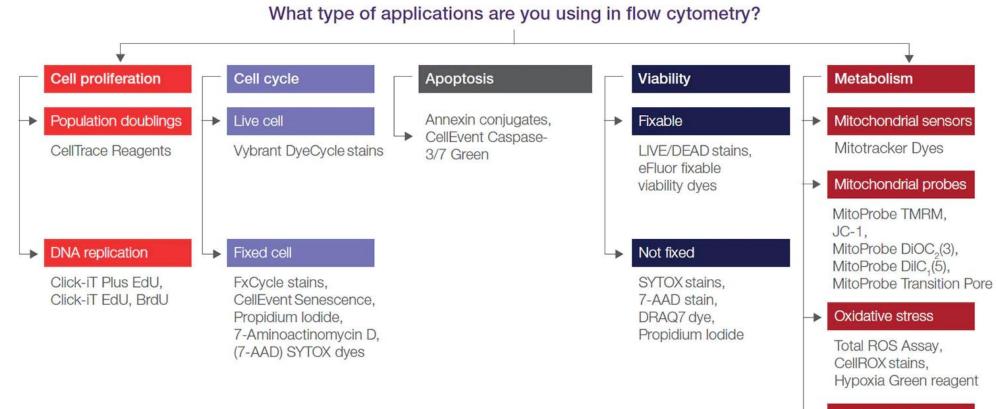


- High level introduction of different dyes/ chemistries and alternative flow cytometry assays
- Understanding of Thermo Fisher flow cytometry resources available (people, online tools, protocols, antibodies, cell health assays, comp beads/ buffers, analysis instrumentation, sorting instrumentation, spectral flow etc..)
- Direction and perhaps even some refreshed motivation for the next flow cytometry steps in your work





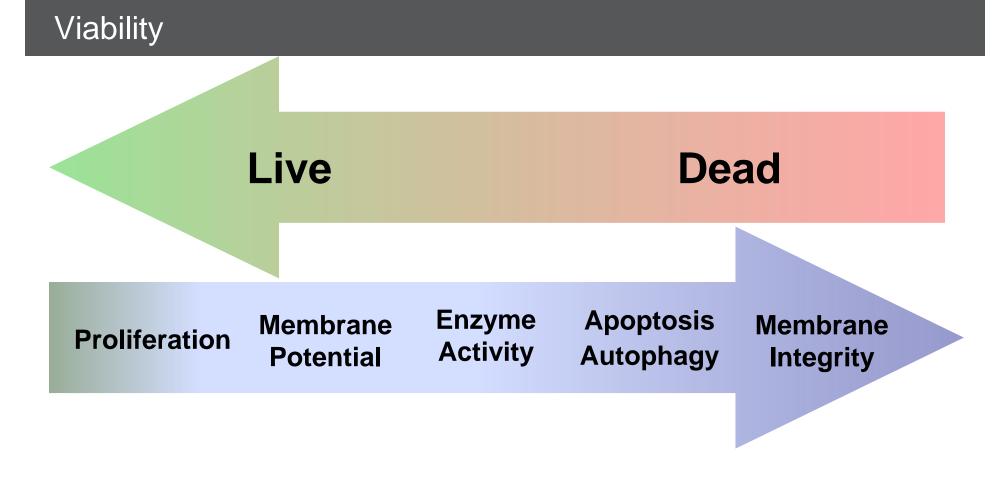




#### Calcium indicators

eBioscience Calcium Sensor Dye eFluor 514, Indo-1 AM





# Cells exist anywhere on a continuum between healthy and dead

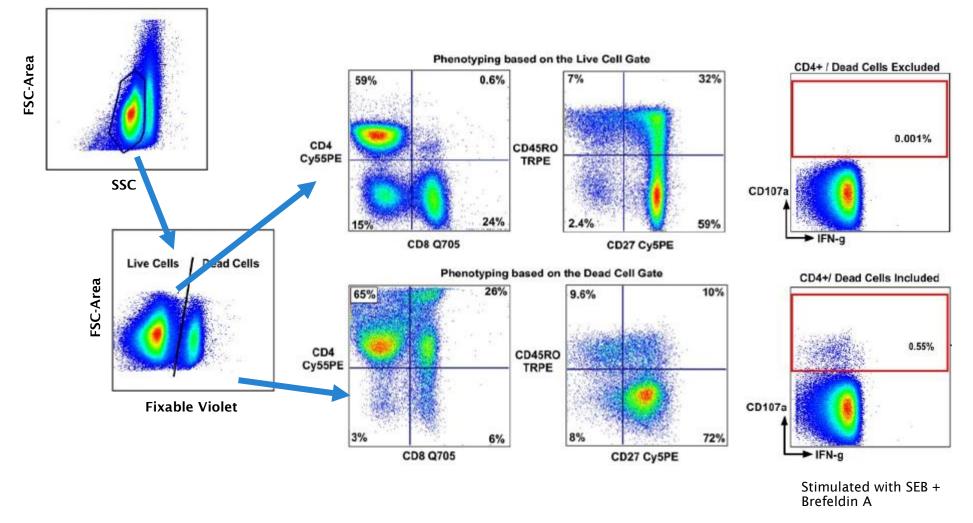


Why is viability important for flow cytometry experiments?

- Dead cells non-specifically bind antibodies, and look like viable cells
- Dead cells bind all kinds of markers
- Potentially lead to erroneous results
- Excluding dead cells improves accuracy of results



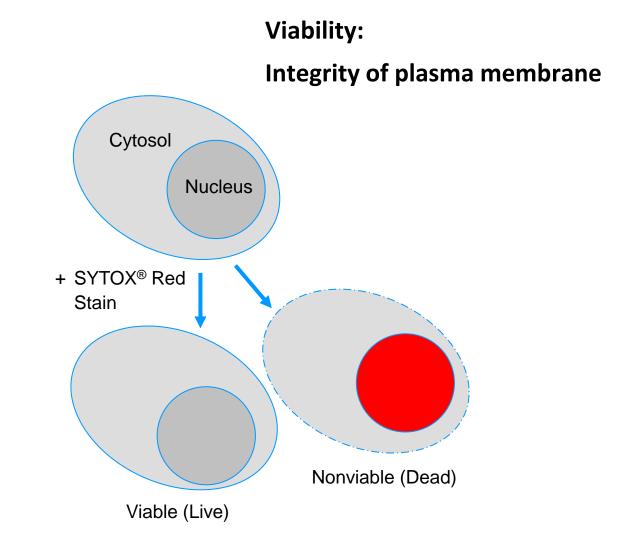
#### Why use a viability indicator?



Perfetto et al. (2006) J Immunol Methods 313:199



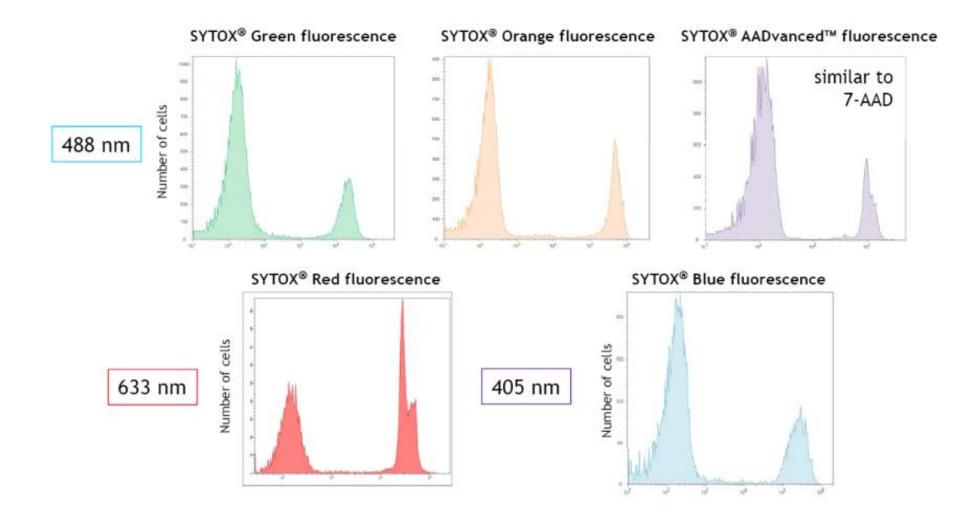
#### Viability: Impermeant nucleic acid-binding dyes





#### Viability: SYTOX<sup>®</sup> Dead Cell Stains

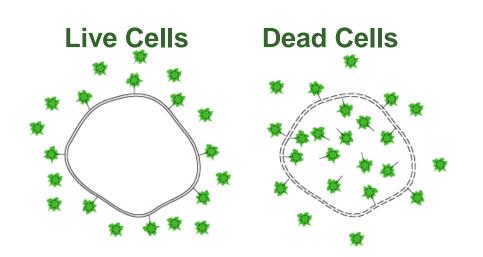
#### Five different colors for flexibility in multicolor panels

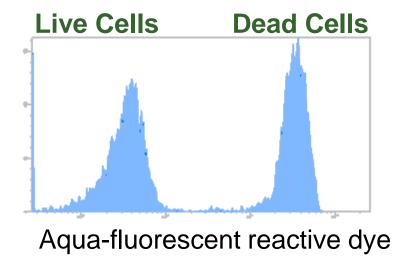




# LIVE/DEAD<sup>™</sup> Fixable Dead Cell Stain Kit

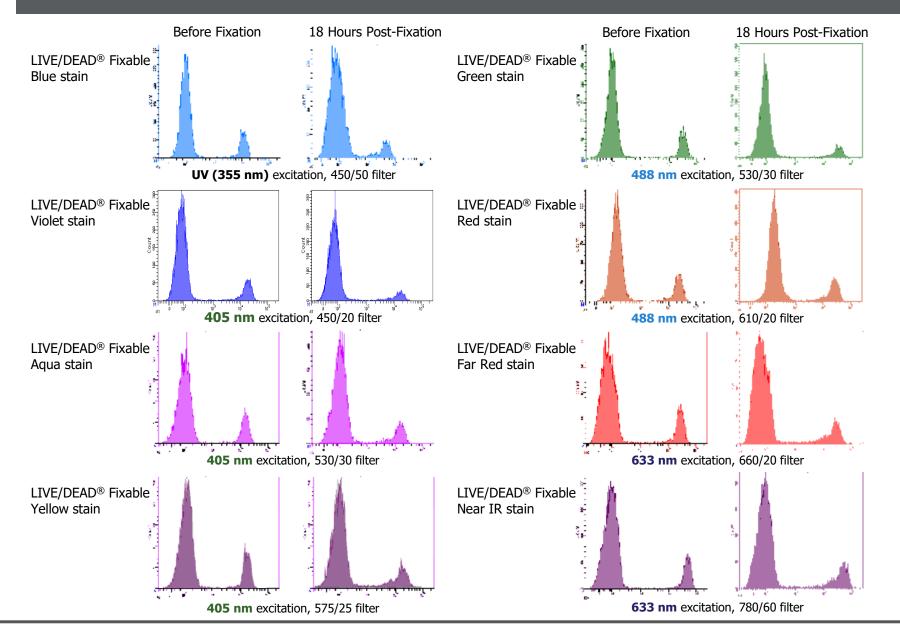
#### eBioscience<sup>™</sup> Fixable Viability Dyes





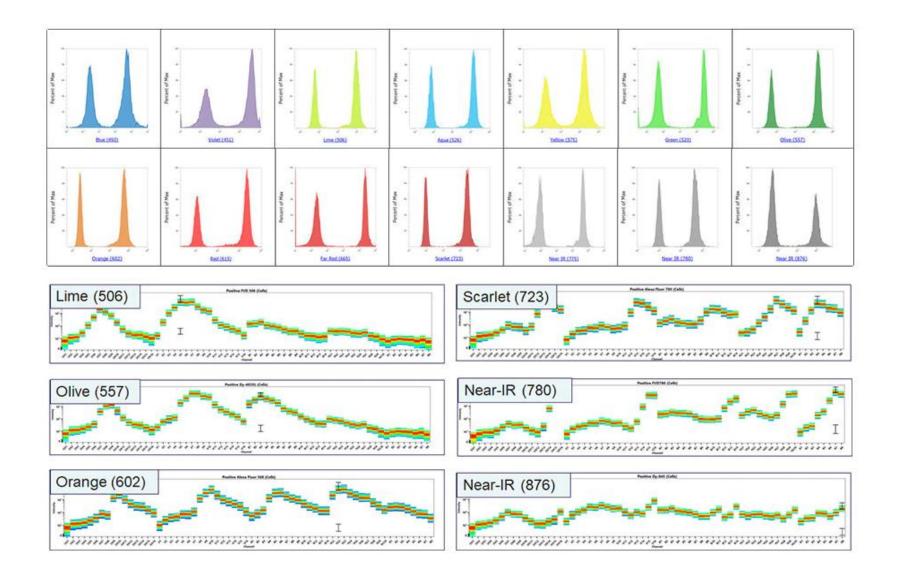


#### Viability: LIVE/DEAD<sup>®</sup> Fixable Dead Cell Stains – amine reactive





#### 14 different dyes now available

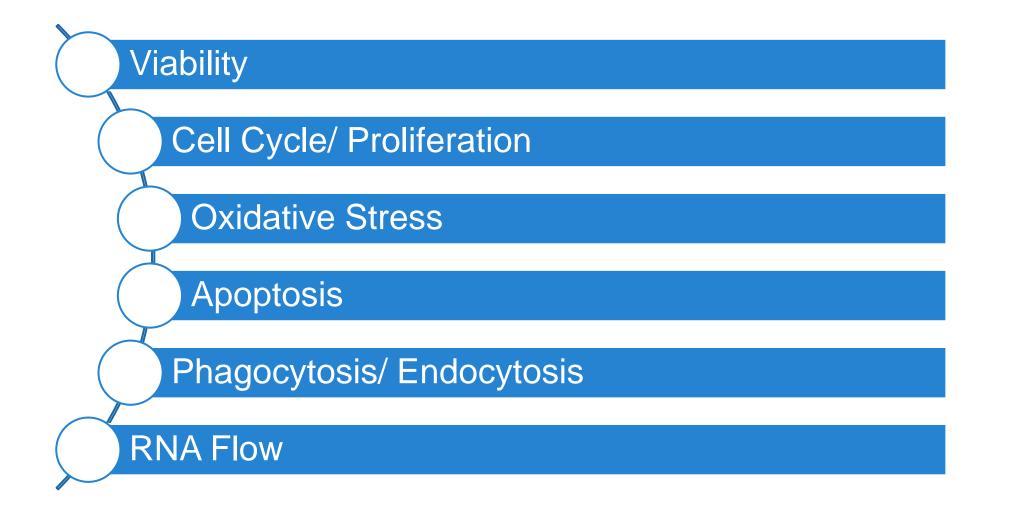




#### Impermeant DNA Dyes

- Add at final step, do not wash out
- Emission is broad, consider for multicolor applications
- Dead/Fixed cells can be used for compensation control
- Amine-reactive Dyes
  - Do not use protein in buffers
  - Live cells have dim fluorescence
  - Use with -aldehyde fixatives
  - Can be used without fixing cells too
  - ArC<sup>™</sup> compensation beads useful

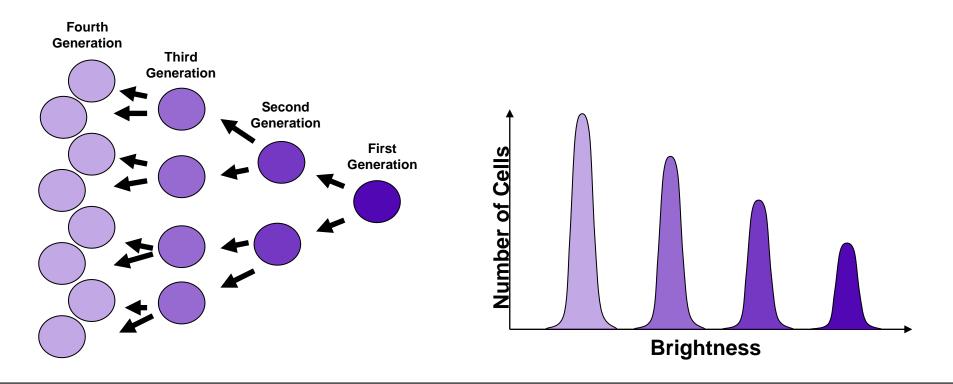






### Cell proliferation analysis by dye dilution

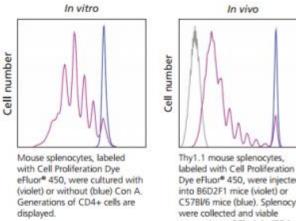
- Cell permeant, cleaved by intracellular esterase giving bright homogenous fluorescence that is well retained compound
- Cell division results in equal partitioning of dye between daughter cells
- Fluorescence of daughter cells is half that of parent cell





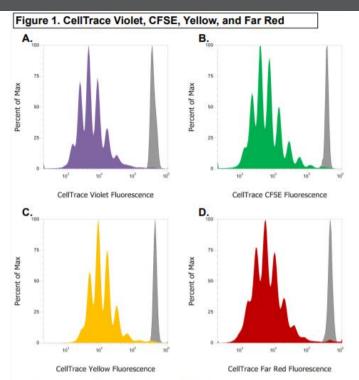
#### Cell proliferation dye

Cell Proliferation Dye eFluor® 450



Thy1.1 mouse splenocytes,
labeled with Cell Proliferation
Dye eFluor® 450, were injected
into B6D2F1 mice (violet) or
C578I/6 mice (blue). Splenocytes
were collected and viable
generations of Thy1.1+/CD4+
cells are displayed. Unlabeled
host cells (Thy1.1-/CD4+) are
shown in grey.

Cell Proliferation Dyes					
Description	Excitation	Emission	Size	Cat. No.	
Violet Laser					
Call Proliferation Due of Lucr® 450	409 nm	450 nm	500 µg	65-0842-85	
Cell Proliferation Dye eFluor® 450	409 nm		4 x 500 µg	65-0842-90	
Blue Laser					
CFSE	488 nm	521 nm	5 x 500 µg	65-0850-84	
Red Laser					
Call Proliferation Due of luor® 670	622.000	670 nm	500 µg	65-0840-85	
Cell Proliferation Dye eFluor® 670	633 nm		4 x 500 µg	65-0840-90	

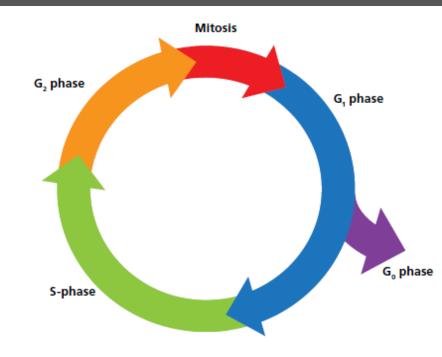


PBMC's were stained with (A) 5 µM CellTrace Violet, (B) 2 µM CellTrace CFSE, (C) 10 µM CellTrace Yellow, and (D) 2 µM CellTrace Far Red. Dynabeads Human T-Activator CD3/CD28 were used for T cell expansion and activation. Samples were incubated in OpTmizer T-cell Expansion Medium at 37th C / 5% CO<sub>2</sub> for 7 days. Samples were analyzed using SYTOX Green or SYTOX Red dead cell stains to gate on live cells and mouse anti-human CD4 Pacific Blue or CD4 FITC were used to gate on proliferating cells. The gray peaks represents unstimulated control cells (parent generation) and the peaks to the left of each gray peak represent individual generations of cells that proliferated during the course of the experiment.

Fluorescent label	CellTrace Blue	CellTrace Violet	CellTrace CFSE		CellTrace Far Red
Laser	UV	405	488	532, 561	633/635
Ex/Em (nm)	355 or 375/410	405/450	495/519	546/579	630/661
Multiplexable	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
20 tests	C34574	C34571	C34570	C34573	C34572
180 tests	C34568	C34557	C34554	C34567	C34564



#### Cell cycle and proliferation by flow



#### Cell cycle analysis

- Go phase: Resting cells have zero growth
- G, phase: Enzyme synthesis is required for DNA replication
- S-phase: DNA replication producing two identical sets of chromosomes
- G, phase: Protein synthesis occurs
- Mitosis: The nucleus and cell divide

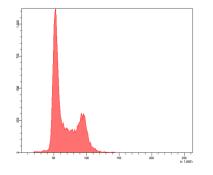
			D
Method	is to Eval	uate Cell	Proliferation
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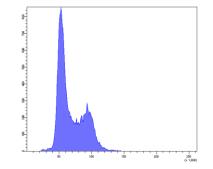
BrdU	Ki-67	PCNA	Proliferation Dyes
Measures cells in S-phase only	Measures proliferating cells at	Measures S-phase but also	Measures generational
	any cell cycle stage except G <sub>o</sub>	includes late G, phase	proliferation
Pulse-labeling common to avoid	BrdU is a subset of Ki-67	Data supports IHC applications	Long-term labeling assay.
cytotoxicity	positive cells	Not as robust for flow cytometry	Does not require fixation
In long-term culture, BrdU can be pulse-labeled and washed out Dividing cells will not incorporate BrdU so toxicity is diluted	Ki-67 and BrdU are used together in both IHC and		Cannot distinguish cell cycle phases of daughter cells



# Cell-permeant nucleic acid dyes

- Dyes which have the ability to penetrate an intact cell membrane to stain nucleic acid
- Used for determining the DNA content of *viable* cells.
- Allows resolution of cell cycle information against the dynamic background of LIVING cells
  - Hoechst dyes (UV ex) dsDNA(A-T)
  - Vybrant<sup>®</sup> DyeCycle<sup>™</sup> Violet stain (UV, 405 ex) dsDNA
  - Vybrant<sup>®</sup> DyeCycle<sup>™</sup> Green stain (488 ex) dsDNA
  - Vybrant<sup>®</sup> DyeCycle<sup>™</sup> Orange stain (488 & 532 ex) dsDNA
  - Vybrant<sup>®</sup> DyeCycle<sup>™</sup> Ruby stain (488–633 ex) dsDNA







• Dyes which penetrate cells with a compromised cell membrane to stain nucleic acids, but do not cross the membranes of live cells

# >Can be used to identify dead cells in a population >Can be used to quantitate DNA content in fixed cells

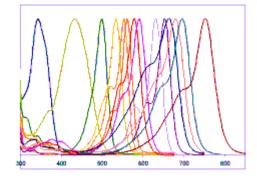
- Propidium Iodide (488 nm ex)
- 7-AAD (488 nm ex)
- SYTOX<sup>®</sup> AADvanced<sup>™</sup> dead cell stain (488 nm ex)
- SYTOX<sup>®</sup> Green dead cell stain (488 nm ex)
- SYTOX<sup>®</sup> Orange dead cell stain (488 /532/561 ex)
- SYTOX<sup>®</sup> Blue dead cell stain (405 nm ex)
- SYTOX<sup>®</sup> Red dead cell stain (633 nm ex)

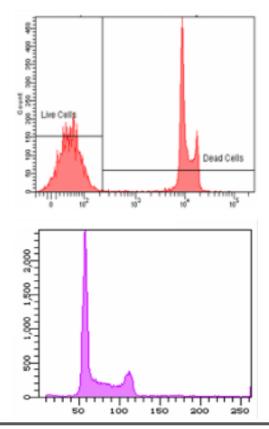


### Which dye to use for cell cycle analysis?

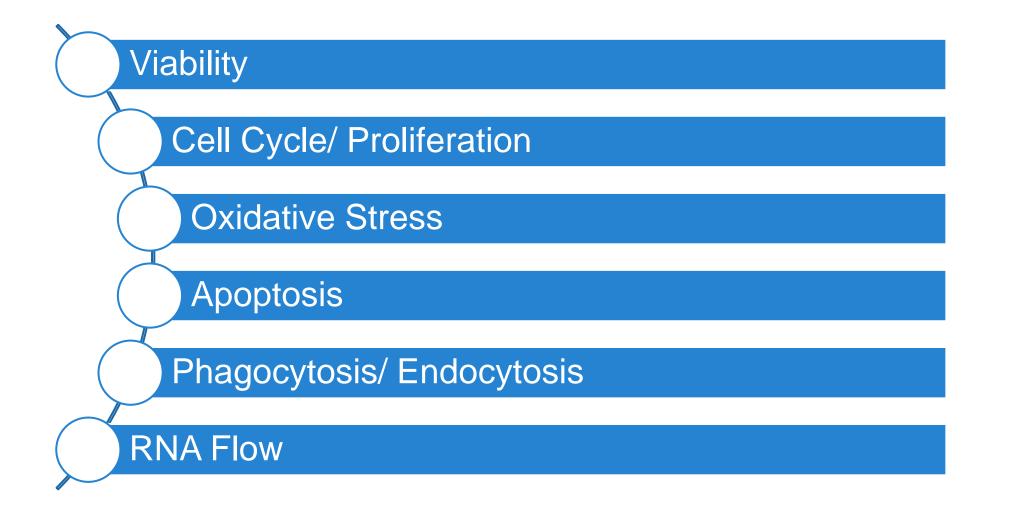
## Things to think about:

- Excitation source
- Emission of dye
- Instrumentation
- Application
- Specificity
- Live DNA content analysis
- Fixed DNA content analysis











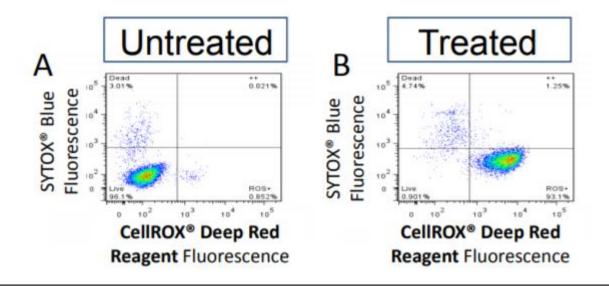
- Oxidative stress imbalanced production of reactive oxygen species (ROS) and the ability of the cell to scavenge them
- ROS react with nucleic acids, proteins and lipids causing cell and tissue damage
- Can be measured using ROS indicators.

	CellROX Deep Red Reagent	CellROX Orange Reagent	CellROX Green Reagent	CM-H <sub>2</sub> -DCFDA
Common filter set	Cy5	RFP	FITC	FITC
Reporter	CellROX Deep Red Reagent	CellROX Orange Reagent	CellROX Green Reagent	H <sub>2</sub> -DCFDA
Ex/Em (nm)	640/665	545/565	485/520	495/527
Cat. No.	C10422	C10443	C10444	C6827

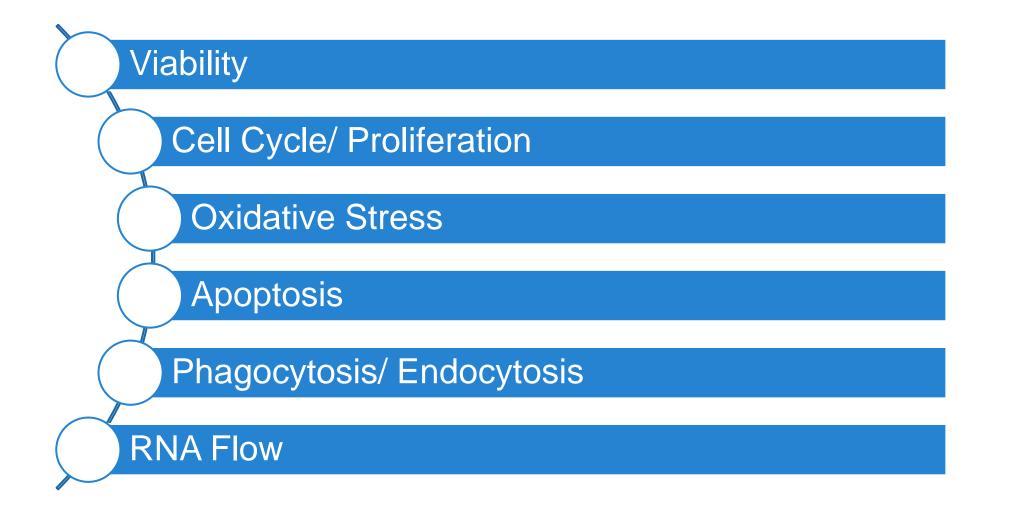


#### Oxidative stress - CellROX

- Fluorogenic probes for measuring generalized oxidative stress in cells
- Detection and quantitation of reactive oxygen species (ROS)
- Live cell compatible (cell permeable)
- Nonfluorescent in a reduced state and fluoresce upon oxidation by ROS
- Some are fixable/detergent resistant for multiplexing
- Reagent can be applied to cells in complete growth media









# Plasma membrane changes

- Annexin V
- Membrane permeability
- Membrane asymmetry

# Nucleus

- Chromatin condensation
- DNA fragmentation

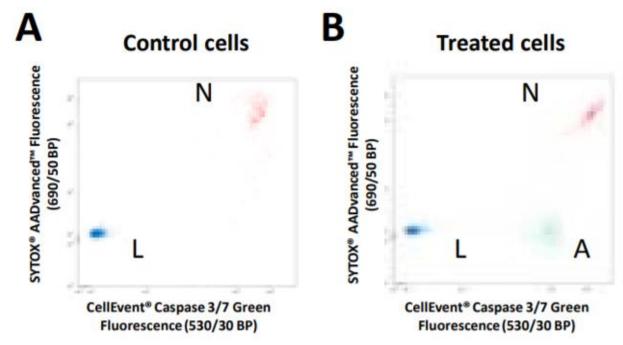
- Mitochondrial membrane changes
  - Membrane potential
  - Mitochondrial transition pore

- Cytoplasm
  - Caspase activity



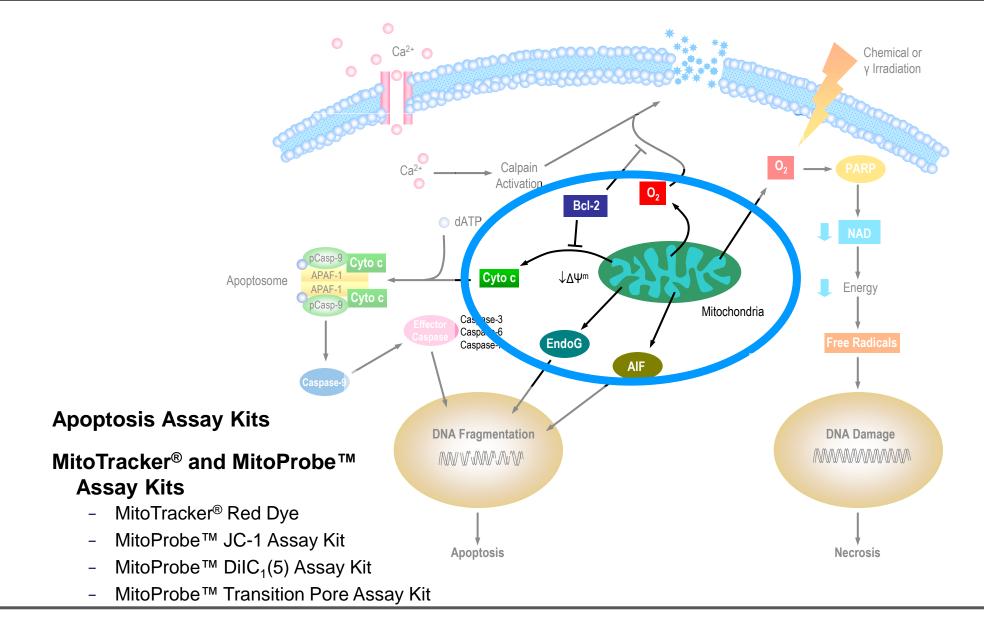
#### Apoptosis - caspase detection

- CellEvent Caspase-3/7 Green Flow Cytometry Assay Kit (<u>C10740</u>, <u>C10427</u>)
  - Detection of activated caspases 3 and 7
  - Cell-permeant reagent conjugated to nucleic acid binding dye
  - Reagent contains caspase-3/7 recognition sequence
  - Fluorescent detection upon cleavage and DNA binding

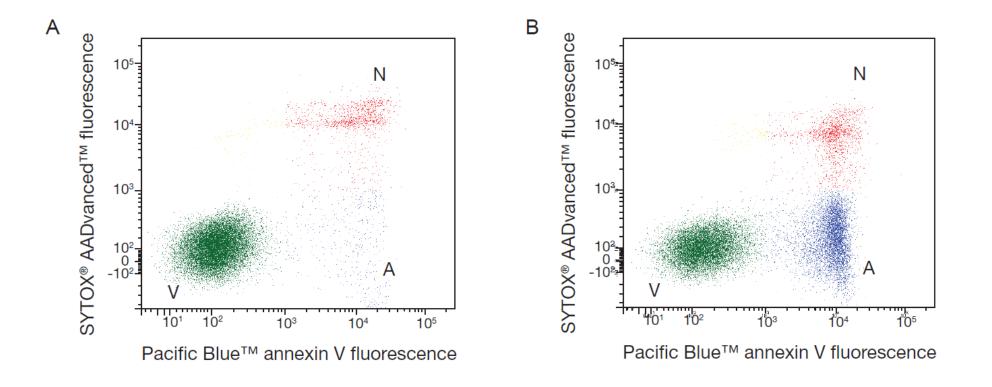




#### MitoProbe<sup>™</sup> Reagents: Detecting mitochondrial changes

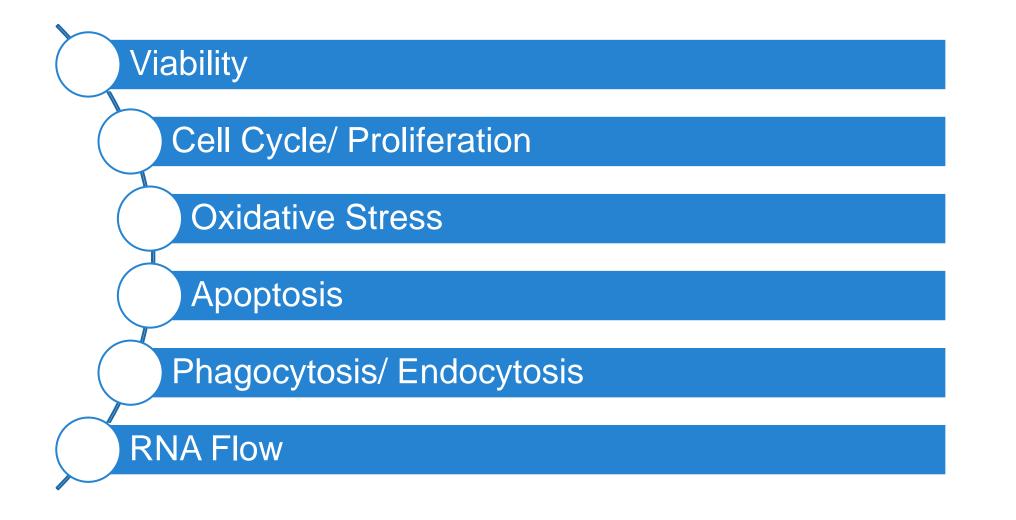






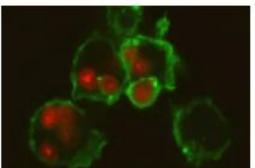
**Figure 1.** Jurkat cells (T-cell leukemia, human) treated with 10 µM camptothecin for four hours (panel B) or untreated control (panel A). Cells were treated with the reagents in the kit and analyzed by flow cytometry using 405 nm and 488 nm excitation. Note that the camptothecin-treated cells have a higher percentage of apoptotic cells (panel B) than the basal level of apoptosis seen in the control cells (panel A). A = apoptotic cells, V = viable cells, N = necrotic cells.



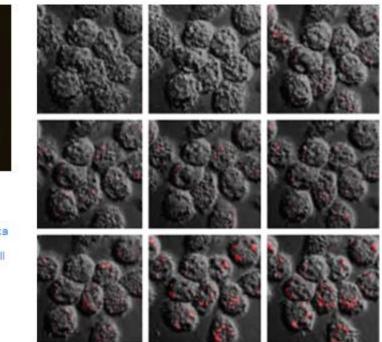




- Cells internalize particulate matter such as microorganisms, and this process is important for immune responses and during the clearance of apoptotic cells
- Tracking phagocytosis using a quench/washbased assay can report on simple uptake, or a pH indicator can be used monitor stages in the pathway.
- pHrodo dyes are essentially non-fluorescent at neutral pH and exhibit increasing signal with a red or green readout respectively as the pH decreases. The increase in fluorescent signal can be used to monitor progression in the phagocytic pathway.



MMM macrophage cells incubated with Zymosan A (S. cerevisiae) BioParticles, Alexa Fluor 594 Conjugate and washed in Live Cell Imaging Solution before imaging.

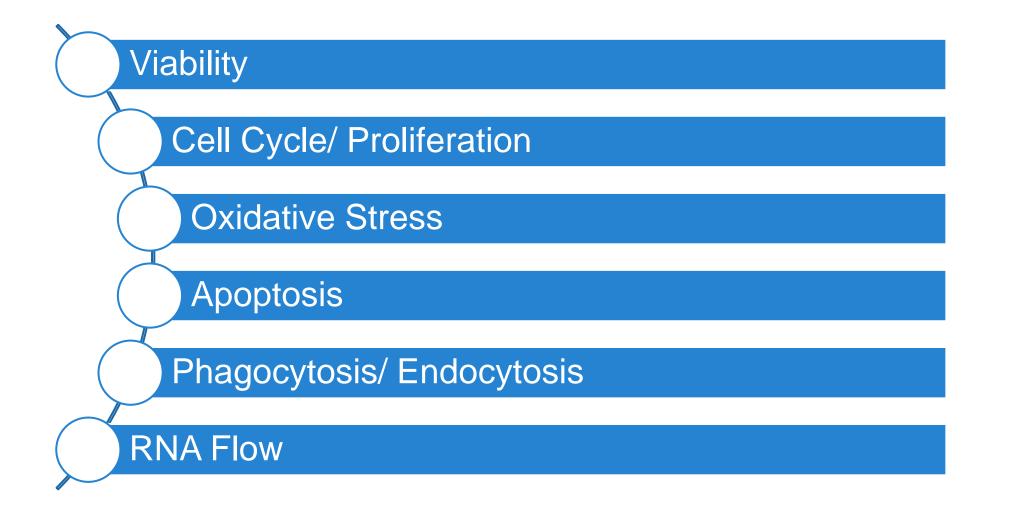




### pHrodo Bioparticles for Phagocytosis

	pHrodo Red <i>E. coli</i> BioParticles Phagocytosis Kit for Flow Cytometry	pHrodo Red Phagocytosis Particle Labeling Kit for Flow Cytometry	pHrodo Green <i>E. coli</i> BioParticles Phagocytosis Kit for Flow Cytometry	pHrodo Green S. aureus Bioparticles Phagocytosis Kit for Flow Cytometry		
Readout	v cytometry					
Range		Monitors phagosome formation				
Vehicle or Method	E. coli	Label your own particles	E. coli	S. aureus		
Common filter set	TRITC		FITC			
Labels	pHrodo Red		pHrodo Green			
Ex/Em (nm)	m (nm) 500/585		509/533			





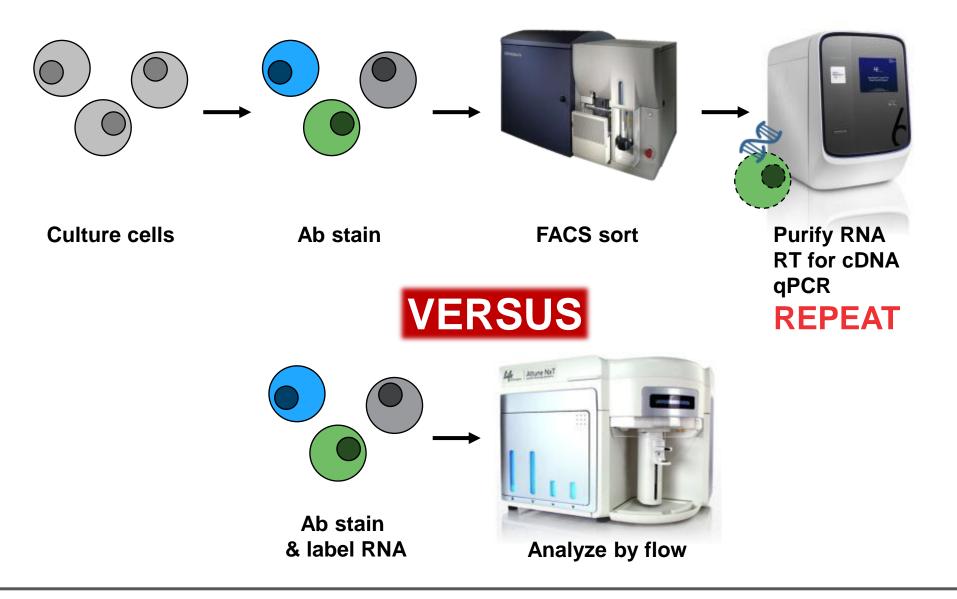




- RNA AND protein expression
   in millions of single cells
- Novel *in situ* hybridization assay: Simultaneous detection of 4 RNA (mRNA, IncRNA, vRNA & miRNA!) using your flow cytometer
- Combine with your antibodies:
   Cell surface and IC proteins

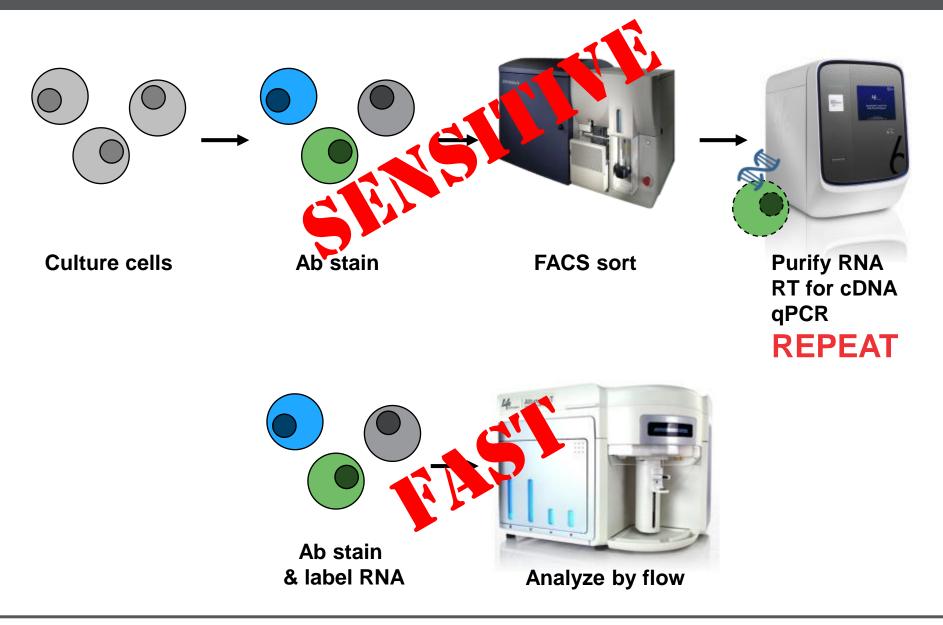


#### Transcript characterization... until now



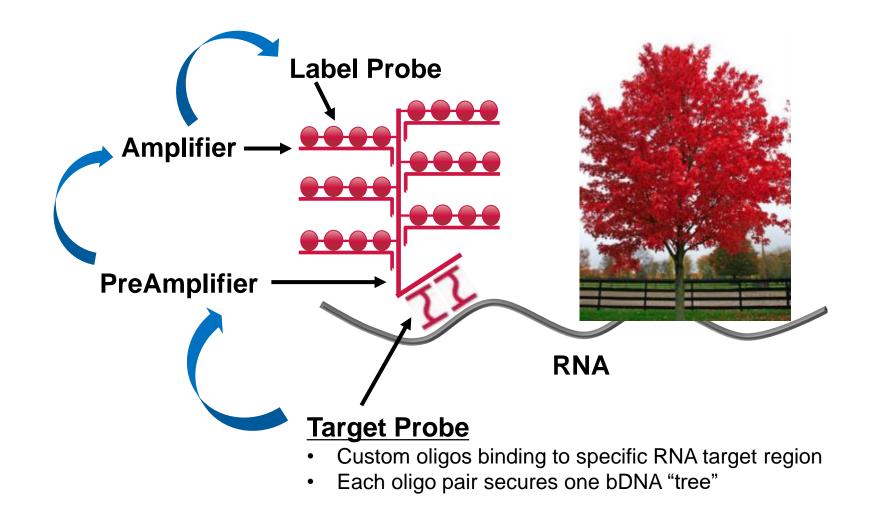


#### Transcript characterization... until now

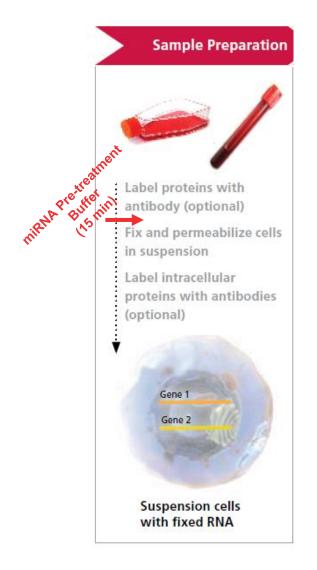




#### Branched DNA: How does it work?

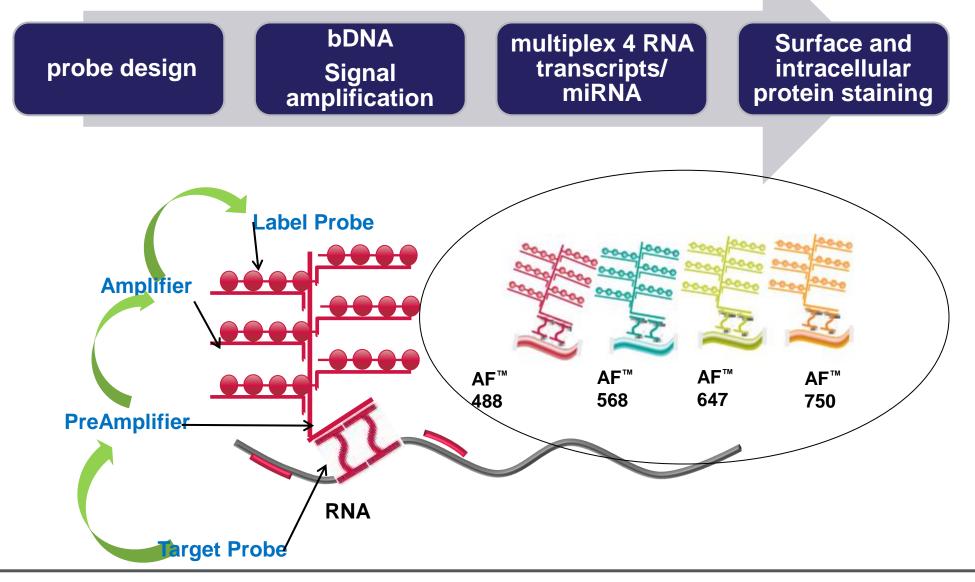








### PrimeFlow RNA assay allows detection of (mi)RNA by Flow





### Gene expression heterogeneity RARE POPULATIONS

Simultaneous transcription + translation studies
 Cytokines

# • Viral immunology

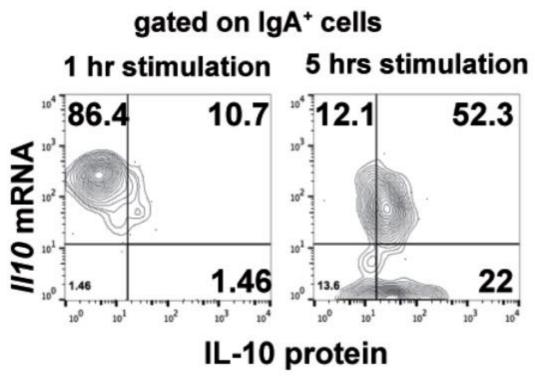
Replication and latency (RARE POPULATIONS!)

## • No antibody? No problem!



IgGA<sup>hi</sup> B cells regulate tumor-infiltrating CD8 T cells

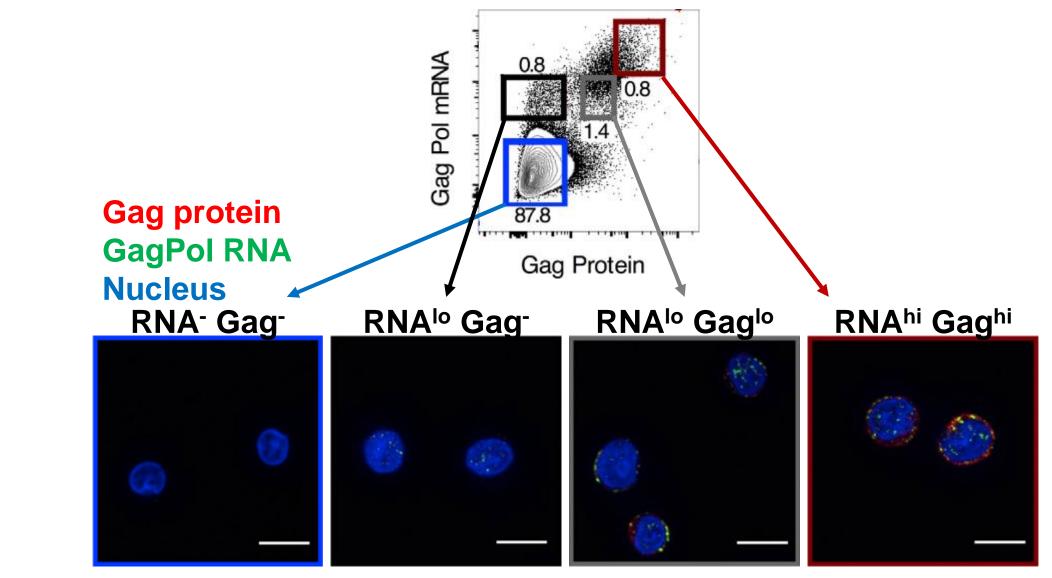
Mouse prostate cancer model



Shalapour et al., 2015, Nature



#### Visualizing HIV protein & RNA

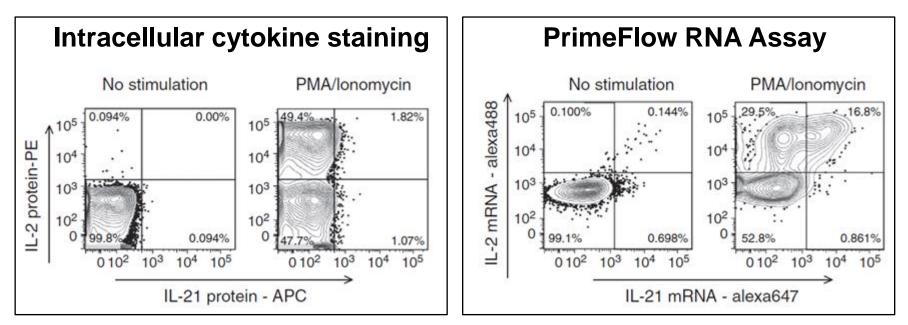


Baxter et al., 2016, Cell Host & Microbe



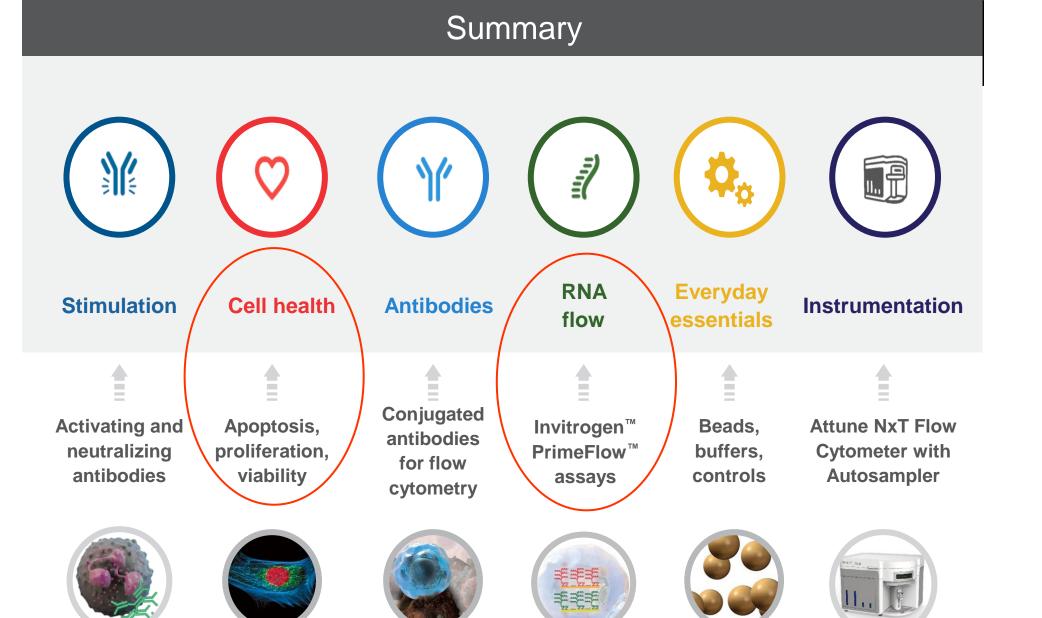
# IL-21 in Idiopathic thrombocytopenic purpura (ITP)

No good Ab for IL-21!



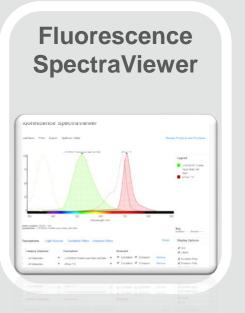
Porichis et al., 2014, Nature Communications







#### Antibody and Flow Cytometry Experimental Resources



Multicolor panel building

Learn more at: thermofisher.com/s pectraviewer



Antibody

Antibody education and purchase

Learn more at: thermofisher.com/fl owantibodies

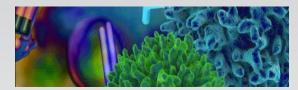
#### Invitrogen™ eBioscience™ reagents



Immunology antibody education and purchase

Learn more at: thermofisher.com/ant ibodies/ebioscience

#### Flow Cytometry Learning Center



Flow Cytometry Learning Center Learn more about flow cytometry applications, techniques, and basic principles.



#### Molecular Probes<sup>™</sup> School of Fluorescence—Flow\_Cytometry Basics

Learn how a flow cytometer works including the fluidics, optics and electronics. This is a free resource to help you get started with flow cytometry, which can be a complex and challenging application.

#### Flow Cytometry Resource Library

Curated collection of scientific application notes, publications, videos, webinars, and scientific posters for flow cytometry.



- Flow Cytometry Application Notes, Scientific Posters, and BioProbes Articles Various applications, providing the conditions and reagents used to achieve the results. Scientific posters presented by our R&D scientists at key conferences. Articles from BioProbes Journal.
- **T Cell Stimulation and Proliferation eLearning Course** Modular, animated, and narrated eLearning course on T cell activation and the methods used to measure T cell function. Knowledge checks and a practical application session.
- Flow Cytometry Educational Videos & Webinars Media for researchers interested in flow cytometry.
- Flow Cytometry Research Tools Fluorescence SpectraViewer, flow cytometry panel design tool, antibodies search tool, mobile apps and more.
- Flow Cytometry Protocols Step-by-step instructions for successful fluorescence-based assays to measure cell proliferation, viability, and vitality using your flow cytometer.
- The Molecular Probes ™ Handbook—A Guide to Fluorescent Probes and Labeling Technologies Extensive references and technical notes. Contains 3,000 technology solutions representing a wide range of biomolecular labeling and detection reagents.





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# **Questions?**

Thank you!

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